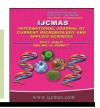


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Heterosis Study for Yield, Water Use Efficiency and its Attributing Traits in Greengram (*Vigna radiata* L. Wilczek)

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ABSTRACT

Keywords

Heterosis, WUE, Abiotic stress, L x T, SLA, SCMR

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Seven lines, four testers and twenty eight F_1 crosses derived through line x tester mating were evaluated to study the magnitude of heterosis for yield, water use efficiency and its attributing traits in order to assess the worth of a cross. The study on extent of heterosis revealed that sixteen hybrids out of twenty eight hybrids recorded high status with maximum score. The hybrid BL 862 × KKM 3 exhibited highest significant negative mid parent heterosis in desirable direction for days to 50 % flowering followed by BL 862 × VGG 4, Hg 19/A × NP 36 and LM 567 × KKM 3. The hybrid BL 866 x KKM 3 exhibited significant MP and BP heterosis for primary branches per plant, where as BL 862 × NP 36 showed significant mid parent heterosis for clusters per plant. BL 862 × GM 8413 (-7.31 %), BL 866 × VGG 4 (-6.21 %) and BL 862 × GM 8413 (-7.35 %), BL 866 × NP 36 (-6.69 %) showed significant negative heterosis over MP and BP for Δ ¹³C value, cross SML 134 × VGG 4 (51.75 %) revealed positive significant heterosis over better parent for SCMR, these may be useful in future breeding programs.

Introduction

Mungbean (Vigna radiata (L) Wilczek) (2n=2x=22). One of the important pulse crops in India as well as in South East Asia. It is primarily a crop of rainy season. However, with the development of early maturing varieties, it has proved to be an ideal crop for spring and summer seasons. Water stress reduces plant growth and yield. However, water stress that exists at the reproductive severely affects grain yield mungbean more than its occurrence at other stages (Thomas et al., 2004). In addition, the time of flowering and maturity was shortened under stress compared to well

conditions. Progress in crop improvement for water use efficiency was rather slow because a concomitant reduction in total biomass while selecting for increase water use efficiency is perhaps the major reason for the lack of success in plant breeding programme (Kumar *et al.*, 1998). Study of heterosis is important for the plant breeder to find out the superior crosses in first generation itself. In addition to this, the magnitude of heterosis provides basis for determining genetic diversity and also serves as guide to the choice of desirable parents. An attempt was, therefore made to know the magnitude of

heterosis over mid parent and better parent for seed yield, water use efficiency and its attributing traits in elite Indian mungbean genotypes (Gwande *et al.*, 2001 and Joseph and Santhoshkumar, 2000).

Materials and Methods

Seven mungbean cultivars based on low Δ^{13} C values were selected as lines (BL-866, BL-862, Hg19/A, LM-567, ML-347, RMG-62 AND SML-143) were crossed with four high yielding genotypes (NP-36, VGG4, GM-8413 and KKM-3) which were selected as testers based on high Δ^{13} C values and were of high yielding in Line × Tester fashion during kharif 2009-2011 at University Agricultural Sciences, Bangalore, to generate a total of twenty eight hybrids. All twenty eight hybrids were sown along with their parents in randomized complete block design with two replications and evaluated during rabi 2010. Each entry was sown in 4m length with 30 x 10 cm spacing in single row. Recommended cultural practices were followed.

Specific leaf area (SLA, cm2/g)

Third fully expanded leaf of the main branch was collected and the leaf area was measured using leaf area meter. Then the leaves were kept in an oven at 70°C for 3 days. The dry weight of the leaf was accurately measured using a sensitive balance. SLA was computed using the formula:

SPAD chlorophyll meter reading

Amongst several leaf characters, leaf thickness and chlorophyll content determines the leaf transmittance characters. Leaf nitrogen content normally influences the leaf chlorophyll content. A device has been developed by Minolta company, New Jersey USA (SPAD-502) which measures the light attenuation at 430nm (the peak wavelength for chlorophyll a and chlorophyll b) and at 750 nm (near infra-red) with no transmittance. The unit value measured by the chlorophyll meter is termed as SCMR (SPAD chlorophyll meter reading) which provides information on the relative amount of leaf chlorophyll. The **SPAD** meter (Soil Plant Analytical Development) is a simple hand held instrument, which operates with DC power of three volts. The third leaf from the apex was selected to record the SCMR. Selected leaf of green gram was clamped avoiding the mid rib region into the sensor head of SPAD meter. A gentle stroke was given to record the SPAD reading and the average of such four strokes was considered. Since mungbean has trifoliate leaf, SCMR was recorded in all leaflets and the average value wad recorded. The SCMR was recorded under normal sunlight between 9.00 am to 4.00 pm.

Mass spectrometric analysis

 $(^{13}C/^{12})$ Carbon isotope ratios C) comparison with the Pee and Dee Belemnite standard were measured using continuous flow isotopic mass spectrometer (IRMS). The IRMS facility consists of flash elemental analyzer (CE-EA 1112), for sequential combustion of biomass samples and open slit interference (coulo 3). Finely powdered dry leaves samples were accurately weighed in the range of 1.0 to 1.2 mg into silver capsules. The crimped capsules with the sample were placed sequentially in the caraousel of the auto sampler. The samples are dropped at specific interval of time along with a pulse of pure O2 in to the oxidation reactor.

The combustion (oxidation) reactor contains chromic oxide and silvered cobaltouscobaltic-oxide heated to 105° C. The biomass is completely oxidized to produce CO_2 , N_2O and H_2O . These gases were swept into the reduction furnace using helium carrier gas. The reduction column contains reduced copper in quartz tubes heated to 680° C. In this reaction, the N_2O is reduced to N_2 and the excess O_2 is absorbed. The resultant gases are then flushed through scrubbers to trap CO_2 and water.

The pure CO₂ and N₂ gas after passing through a GC column (5° A molecular sieve) and a thermal conductivity detector (TCD) into the ion source of IRMS. At the source, CO₂ is ionized by electron impacts ionization to produce molecular radicals (CO+). When accelerated radicals pass through a strong magnetic field it is deflected with the radius of deflection being proportional to the molecular mass of the radicals. These deflecting ¹² CO₂ and ¹³ CO₂ are collected by the Faraday cups and the signal is amplified and transmitted to the computer and displayed. Based on fractionation (isotopic composition with respect to PDB), the ¹³C discrimination (Δ^{13} C) in the plant sample was computed as follows and expressed as parts per thousand (%) or per mil.

$$\Delta^{13} C (\%) = \frac{\delta_a^{13} C - \delta_p^{13} C}{1 + \delta_p^{13} C/1000}$$

All plants in a cross were tagged and observations were recorded on individual plant basis. The observations on water use efficiency, yield and its attributes *viz.*, Δ13C, SLA (Specific leaf area), SCMR (SPAD chlorophyll meter reading), days to 50% flowering, plant height, primary branches per plant, clusters per plant, pods per cluster, pods per plant, pod yield per plant, pod length, seeds per pod, seed yield per plant, threshing percentage were recorded and subjected to statistical analysis. Improving water use

efficiency (WUE) becomes quite relevant among the number of traits suggested for crop improvement under such conditions. The yield model proposed by Passioura (1986):

Seed yield = Water use efficiency (WUE) \times Total transpiration (T) \times Harvest index (HI)

The mean values of the data recorded for water use efficiency, seed yield per plant and their attributing characters in mungbean in each entry of both the replications were first subjected to Analysis of Variance as per the methods outlined by Panse and Sukhatme (1967) using mean values of randomly selected plants. All sources of variation were tested against error for significance by comparing calculated 'F' value with table 'F' value at 1 per cent and 5 per cent probability levels.

The treatments mean value for each trait was used for the estimation of heterosis. The per cent heterosis of all F_1 crosses over their better parent (BP) and mid parent (MP) were computed by the method suggested by Turner (1953) and Hayes *et al.*, (1955).

Overall gca status of parents, sca status and heterotic status of crosses

Since yield is associated with several other characters, positively with some and negatively with others, it is necessary to know the overall status of the parents / hybrids considering gca effects / sca effects for all the characters simultaneously. The overall status of a parent or a cross with respect to gca or sca effects was determined as per the method of Arunachalam and Bandyopadhyay (1979) with slight modification as suggested by Mohan Rao (2001). The modified procedure is described as under.

The estimates of general combining ability effects of parents, sca effects and standard

heterosis of hybrids were ranked by giving the highest rank for the parent or the cross which manifested the highest gca / sca effects and mid parent heterosis respectively in desirable direction. The lowest rank was given for parent or the cross with the lowest gca / sca effects and mid parent heterosis for a character, respectively. This was repeated for each character except days to 50 per cent flowering, for which the ranking was given in reverse order. The ranks obtained by the parent / hybrid were summed up across all the characters to arrive at a total score for each of the parent / cross. Further, the mean of the total scores of all the genotypes (parents and hybrids) was computed which was used as the final norm to ascertain the status of a parent or a hybrid with respect to gca / sca effects and mid parent heterosis. The parent / hybrid whose total rank exceeded the final norm were given high (H) overall gca / sca / heterosis status, respectively. On the other hand, the parents or the cross, whose total rank was less than the final norm were given low (L) overall gca / sca / heterosis status, respectively.

Accordingly the crosses were grouped into different category viz., High \times High (HH), High \times Low (HL), Low \times High (LH) and Low \times Low (LL) based on overall gca status of their parents. The overall sca status of crosses viz., High or Low was also mentioned under each category to draw the inference.

Results and Discussion

Heterosis is usually expressed in the form of increased yield in F_1 's which in turn is dependent on the contribution of many component characters. All the component characters of yield, water use efficiency and its attributing traits were studied for heterosis manifestation in order to assess the worth of a cross. When significant heterosis over better parent is observed in majority of the crosses

for any trait, it indicates involvement of nonadditive gene action in the genetic control of that trait. Assuming that epistasis is absent, the cause of heterosis can only be attributed to dominance gene action. The inference drawn about gene action based on heterosis for different characters is presented below based on this assumption.

The extent of heterosis over mid parent (MP) and better parent (BP) expressed by 28 hybrids in respect of grain yield, WUE and its attributing traits have been tabulated in table 1 and briefly presented below trait wise.

The negative heterosis is desirable for days to 50% flowering. Among 28 crosses, the cross BL 862 × KKM 3 exhibited highest significant negative mid parent heterosis (-31.63 %) for days to 50 per cent flowering in desirable direction followed by BL 862 × VGG 4 (-28.57 %), Hg 19/A × NP 36 (-23.16 %) and LM $567 \times KKM \ 3 \ (-22.58 \ \%)$. Of the above crosses, BL 862 × VGG 4 exhibited maximum significant negative heterosis (-32.04 %) over better parent followed by LM $567 \times KKM \ 3 \ (-22.58 \ \%)$, and BL $866 \times$ VGG 4 (-17.20 %). These findings are in agreement with Naidu and Styanarayan (1993). The cross BL 866 × KKM 3 showed significant positive heterosis over mid parent (114.29 %) for primary branches per plant. Whereas, BL 866 × KKM 3 recorded highest significant positive heterosis (114.29 %) over better parent. These findings are in agreement Khattak et al., (2002e).

The cross BL $862 \times NP$ 36 showed significant positive heterosis over mid parent (87.61%) for clusters per plant. These findings are in agreement Khattak *et al.*, (2002e).

Four crosses revealed significant heterosis over mid parent for pods per cluster. However only one cross ML $347 \times GM$ 8413 showed significant positive heterosis over mid parent

(72.22 %). Whereas, four crosses revealed significant heterosis over better parent these findings are in agreement with Khattak *et al.*, (2002e). However none of cross showed positive heterosis over better parent.

12 crosses revealed significant heterosis over mid parent for pods per plant. Only one cross RMG 62 × VGG 4 showed significant positive heterosis over mid parent (25.23 %) (Khattak *et al.*, 2002e).

Whereas, 11 crosses revealed significant heterosis over better parent among that cross Hg $19/A \times GM$ 8413 recorded highest significant negative heterosis (54.10 %) over better parent.

19 crosses revealed significant heterosis over mid parent for seeds per pod. Only one cross BL $866 \times NP$ 36 showed significant positive heterosis over mid parent (19.15 %).

Whereas, 20 hybrids revealed significant heterosis over better parent among that none of hybrid revealed positive heterosis over better parent (Khattak *et al.*, 2002e and Joseph and Santhoshkumar, 2000).

Four crosses revealed significant heterosis over mid parent for pod length (cm). Only one cross SML $134 \times VGG$ 4 showed significant positive heterosis over mid parent (27.83 %). Whereas, seven hybrids revealed significant heterosis over better parent among that only one cross revealed positive heterosis over better parent (27.08).

15 crosses revealed significant heterosis over mid parent pod yield per plant (g). None of the crosses showed significant positive heterosis over mid parent. Whereas, 18 crosses revealed significant heterosis over better parent none of the crosses revealed positive heterosis over better parent. Eight crosses revealed significant heterosis over mid parent for seed yield per plant (g). None

of the crosses showed significant positive heterosis over mid parent. Whereas, 19 crosses revealed significant heterosis over better parent none of the crosses revealed positive heterosis over better parent (Dethe and Patil, 2008 and Khattak *et al.*, 2002e).

Four crosses revealed significant heterosis over mid parent for threshing percentage. Out of that three crosses showed significant positive heterosis over mid parent *viz.*, RMG 62 × KKM 3 (25.97 %), followed by RMG 62 × NP 36 (23.30 %) and LM 567 × VGG 4 (17.16 %). Whereas, five crosses revealed significant heterosis over better parent out of that two crosses revealed positive heterosis over better parent *viz.*, RMG 62 × KKM 3 (24.05 %), followed by RMG 62 × NP 36 (21.95 %).

10 crosses revealed significant heterosis over mid parent for test weight (g). Out of that 4 crosses showed significant positive heterosis over mid parent viz., BL 866 × KKM 3 (21.83 %), followed by BL 862 × GM 8413 (13.30 %), BL 866 × NP 36 (8.50 %) and LM 567 × GM 8413 (8.05 %).

Whereas, ten crosses revealed significant heterosis over better parent out of that 2 crosses revealed positive heterosis over better parent viz., BL 866 × KKM 3(15.86 %), followed by BL 862 × GM 8413 (12.30%).

Three crosses revealed significant heterosis over mid parent for Δ^{13} C value. Out of that two crosses showed significant negative heterosis over mid parent viz., BL 862 × GM 8413 (-7.31 %), followed by BL 866 × VGG 4 (-6.21 %).

Whereas, three crosses revealed significant heterosis over better parent out of that two crosses revealed negative heterosis over better parent viz., BL 862 × GM 8413 (-7.35 %) followed by BL 866 × NP 36 (-6.69 %).

Table.1 Estimates of mid parent and better parent (BP) heterosis in hybrids for growth, yield and WUE related traits in mungbean

Hybrids		flowering (c		Plant height (cm) Primary branches per pla Heterosis Heterosis		nches per plant	Clusters per plant		Pods per cluster		Pods per plant	
Hybrids	Hete					Heterosis		Heterosis		Heterosis		Heterosis
	MP	BP	MP	BP	MP	BP	MP	BP	MP	BP	MP	BP
RMG 62 × GM8413	4.49	2.20	4.54	3.80	12.50	0.00	7.56	-4.48	18.75	5.56	17.39	-3.57
RMG 62 × KKM 3	-5.56 *	-8.60 **	12.53	4.37	37.50	22.22	23.97	11.94	-29.73	-31.58	-11.32	-16.07
RMG 62 × NP 36	3.41	2.25	-13.31	-20.24 **	-22.22	-22.22	-14.53	-25.37	17.95	9.52	6.67	0.00
RMG 62 × VGG 4	-7.78 **	-10.75 **	34.34 **	12.68	25.00	11.11	36.09	17.91	-12.20	-21.74	25.23 *	19.64
BL 866 × GM 8413	-9.71 **	-13.19 **	-18.96 *	-22.43 *	14.29	14.29	-20.14	-25.86	37.93	33.33	14.29	7.32
BL 866 × KKM 3	-7.34 **	-11.83 **	19.49 *	16.41	114.29 **	114.29 **	36.18	28.67	-17.65	-26.32	-3.30	-12.00
BL 866 × NP 36	-1.73	-4.49	10.84	-2.60	-12.50	-22.22	35.50	23.56	-50.00 *	-57.14 *	-40.00 **	-44.90 **
BL 866 × VGG 4	-12.99 **	-17.20 **	-5.26	-17.03	0.00	0.00	0.18	-9.39	-36.84	-47.83 *	-30.43 *	-37.25 **
ML 347 × GM 8413	-7.94 **	-11.22 **	1.33	-7.29	6.67	0.00	-30.69	-32.69	72.22 **	40.91	22.73	3.85
ML 347 × KKM 3	-13.09 **	-15.31 **	30.13 **	27.35 **	20.00	12.50	61.75	54.26	-31.71	-36.36	9.80	7.69
ML 347 × NP 36	-10.16 **	-14.29 **	13.74	-4.02	-17.65	-22.22	-4.04	-5.00	-11.63	-13.64	-16.83	-19.23
ML 347 × VGG 4	-13.09 **	-15.31 **	31.83 **	20.48	46.67	37.50	-8.26	-8.35	15.56	13.04	2.91	1.92
LM 567 × GM 8413	-7.61 **	-8.60 **	-21.25 *	-28.00 **	-28.57	-28.57	23.33	15.38	-35.48	-41.18	-30.67 *	-33.33
LM 567 × KKM 3	-22.58 **	-22.58 **	-28.74 **	-30.33 **	-14.29	-14.29	23.06	13.15	-33.33	-36.84	-25.84 *	-34.00 *
LM 567 × NP 36	-3.30	-5.38	-39.09 **	-48.64 **	0.00	-11.11	-9.13	-13.40	-15.79	-23.81	-11.36	-20.41
LM 567 × VGG 4	0.00	0.00	53.63 **	40.52 **	42.86	42.86	85.38 *	78.21	-60.00 **	-65.22 **	-35.56 **	-43.14 **
SML 134 × GM 8413	-5.21 *	-9.90 **	-23.77 **	-27.86 **	20.00	12.50	8.16	5.77	-14.29	-28.57	-10.11	-24.53

Table.1 (Continued)

1	2	3	4	5	6	7	8	9	10	11	12	13
SML 134 × KKM 3	-5.15 *	-8.91 **	-13.96	-15.20	6.67	0.00	-13.21	-16.67	-20.00	-23.81	-30.10 **	-32.08 *
SML 134 × NP 36	-1.05	-6.93 *	-15.65 *	-26.63 **	5.88	0.00	-19.76	-20.00	19.05	19.05	-1.96	-5.66
SML 134 × VGG 4	-5.15 *	-8.91 **	19.35 *	5.60	-6.67	-12.50	-6.88	-7.44	22.73	17.39	15.38	13.21
BL 862 × GM8413	-12.37 **	-17.48 **	-35.34 **	-38.57 **	7.69	0.00	38.48	38.08	-2.86	-19.05	-8.70	-25.00 *
BL 862 × KKM 3	-31.63 **	-34.95 **	-31.90 **	-33.14 **	38.46	28.57	-1.42	-3.52	-35.00	-38.10	-33.96 **	-37.50 **
BL 862 × NP 36	-13.02 **	-18.93 **	-33.56 **	-42.01 **	6.67	-11.11	87.61 *	84.53	-28.57	-28.57	-4.76	-10.71
BL 862 × VGG 4	-28.57 **	-32.04 **	-27.43 **	-36.02 **	7.69	0.00	14.09	11.22	-36.36	-39.13	-30.84 **	-33.93 **
Hg 19/A × GM 8413	-8.38 **	-9.89 **	-46.12 **	-49.49 **	14.29	14.29	-48.53	-58.33 *	14.29	14.29	-42.27 **	-54.10 **
Hg 19/A × KKM 3	-9.39 **	-11.83 **	-21.25 **	-30.75 **	14.29	14.29	19.57	-1.79	-21.21	-31.58	-9.91	-18.03
Hg 19/A × NP 36	-23.16 **	-23.60 **	-28.13 **	-30.05 **	-25.00	-33.33	-15.37	-32.50	-14.29	-28.57	-21.82 *	-29.51 **
Hg 19/A × VGG 4	-8.29 **	-10.75 **	-16.00	-32.75 **	42.86	42.86	11.95	-11.31	-45.95 *	-56.52 **	-37.50 **	-42.62 **
S.Em. ±	1.20	1.39	2.64	3.05	0.87	1.01	0.95	1.10	2.10	2.42	2.80	3.24

^{*}Significant at P= 0.05 level **Significant at P= 0.01 level

Table.1 (Continued)

	Seeds per pod Heterosis		Pod length (cm) Heterosis		Pod yield per plant(g)		Seed yield per plant (g)		Threshing %		Test weight (g)	
Hybrids					Hete	rosis	Heterosis		Heterosis		Heterosis	
	MP	BP	MP	BP	MP	BP	MP	BP	MP	BP	MP	BP
RMG 62 × GM8413	-9.52	-9.52	-1.27	-13.38	-19.55 *	-31.39 **	-12.74	-25.39 *	8.61	8.31	-4.70	-8.47 *
RMG 62 × KKM 3	-20.18 *	-23.65 *	-4.03	-12.15	-31.01 **	-37.52 **	-13.51	-22.72 *	25.97 **	24.05 *	-8.51 *	-8.83 *
RMG 62 × NP 36	-13.95	-15.91	-12.91	-21.15 *	-28.00 **	-32.37 **	-11.54	-15.98	23.30 **	21.95 *	-8.08 *	-13.89 **
RMG 62 × VGG 4	-10.60	-12.64	-5.55	-15.62	-22.32 *	-29.17 **	-16.78	-23.10 *	6.62	5.14	-17.65 **	-18.33 **
BL 866 × GM 8413	-32.22 **	-37.64 **	13.99	0.00	-18.02	-28.54 **	-26.74 *	-34.44 **	-11.27	-13.89	-5.24	-5.88
BL 866 × KKM 3	-12.29	-15.80	15.88	6.08	-0.80	-8.02	6.35	-0.28	7.96	6.67	21.83 **	15.86 **
BL 866 × NP 36	19.15 *	-24.00 **	-15.46	-23.46 *	-37.49 **	-39.82 **	-33.70 **	-33.72 **	5.56	1.63	8.50 *	6.42
BL 866 × VGG 4	-32.98 **	-37.00 **	-14.77	-23.85 *	-36.85 **	-41.04 **	-46.73 **	-48.24 **	-16.25 *	-19.60 *	1.91	-1.98
ML 347 × GM 8413	-22.18 *	-25.57 **	1.02	-7.39	8.09	-6.91	-0.95	-14.59	-8.47	-8.61	3.14	-3.47
ML 347 × KKM 3	-32.04 **	-32.04 **	-10.77	-14.44	-8.69	-16.42	-16.26	-24.51 *	-7.42	-8.95	2.84	-7.59
ML 347 × NP 36	-24.44 **	-26.09 **	-7.22	-12.06	-16.66	-20.84 *	-15.74	-19.20	1.52	0.55	-0.75	-4.77
ML 347 × VGG 4	-47.78 **	-48.91 **	-16.18	-21.67	-2.34	-9.99	-5.32	-11.69	-3.46	-4.68	-9.80 *	-18.08 **
LM 567 × GM 8413	-19.30 *	-21.14 *	14.19	9.07	-23.93 *	-25.95 *	-20.28	-20.42	4.61	2.03	8.05 *	4.05
LM 567 × KKM 3	-36.80 **	-38.17 **	-26.10 *	-32.50 *	-22.42 *	-25.58 *	-22.13	-25.81 *	0.09	-0.62	-11.17 **	-11.72 **
LM 567 × NP 36	-30.45 **	-30.45 **	8.58	0.28	-19.88	-25.97 *	-17.36	-25.95 *	3.40	0.04	-4.24	-10.06 *
LM 567 × VGG 4	-31.86 **	-31.86 **	8.14	1.27	-31.31 **	-34.58 **	-19.40	-25.84 *	17.16 *	13.03	0.98	0.42
SML 134 × GM 8413	-5.85	-10.33	5.82	4.25	-28.51 **	-34.42 **	-20.60	-27.47 *	10.41	9.82	1.49	-4.03
SML 134 × KKM 3	-14.29	-21.74 *	-9.90	-12.78	-44.94 **	-46.05 **	-40.56 **	-43.04 **	7.85	5.37	-4.70	-5.91
SML 134 × NP 36	-24.39 *	-29.55 **	-0.73	-2.75	-7.91	-9.66	-12.46	-14.45	-4.41	-4.69	3.13	-4.84
SML 134 × VGG 4	-22.29 *	-27.59 **	27.83 *	27.08 *	9.61	8.20	7.58	6.92	-1.82	-2.40	1.38	-1.08
BL 862 × GM8413	-16.67	-16.67	11.65	6.58	-9.02	-18.02	-19.07	-30.73 **	-11.08	-16.59 *	13.30 **	12.30 **
BL 862 × KKM 3	-33.82 **	-36.70 **	-7.36	-7.41	-21.48 *	-24.54 *	-33.38 **	-40.41 **	-14.55	-21.18 **	1.14	-2.34
BL 862 × NP 36	0.00	-2.27	0.33	-0.83	-12.98	-13.03	-16.45	-20.55 *	-5.32	-10.50	3.99	0.44
BL 862 × VGG 4	-32.88 **	-34.41 **	-8.18	-10.57	-39.60 **	-41.53 **	-38.81 **	-43.39 **	1.61	-3.68	-11.93 **	-13.98 **
Hg 19/A × GM 8413	-38.10 **	-38.10 **	-33.24 **	-38.78 **	-12.26	-20.71	-6.88	-17.41	6.51	3.78	-6.08	-6.57
Hg 19/A × KKM 3	-35.27 **	-38.09 **	-13.30	-16.84	-14.83	-17.89	-17.04	-22.94 *	-2.94	-7.07	-1.36	-5.10
Hg 19/A × NP 36	-33.91 **	-35.41 **	-36.32 **	-39.63 **	-22.46 *	-22.67 *	-32.81 **	-33.47 **	-13.11	-14.65	-4.70	-7.61
Hg 19/A × VGG 4	6.98	4.55	0.05	-6.46	-25.50 *	-27.65 *	-31.98 **	-34.55 **	-9.46	-10.80	4.06	1.27
S.Em. ±	0.93	1.08	0.57	0.66	1.11	1.29	0.70	0.81	4.57	5.28	0.12	0.14

^{*}Significant at P= 0.05 level **Significant at P= 0.01 level

Table.1 (Continued)

	Δ^1	¹³ C	SLA(cı	$n^2/g)$	SCMR		
Hybrids	Hete	erosis	Heter	osis	Hete	erosis	
	MP	BP	MP	BP	MP	BP	
RMG 62 × GM8413	-1.84	-3.34	0.34	-4.71	9.30	3.10	
RMG 62 × KKM 3	-0.84	-2.07	-12.24	-21.25	-6.33	-13.85	
RMG 62 × NP 36	-2.20	-3.16	-1.06	-6.41	-10.25	-15.98	
RMG 62 × VGG 4	-1.71	-2.68	-24.43	-24.55	15.81	11.90	
BL 866 × GM 8413	-4.93	-5.35	38.10	25.44	53.63 *	30.33	
BL 866 × KKM 3	-0.23	-2.53	-3.47	-23.57	53.67 *	-1.61	
BL 866 × NP 36	-4.72	-6.69 *	21.87	11.12	10.29	4.82	
BL 866 × VGG 4	-6.21 *	-6.32	2.44	-11.04	-13.89	-20.66	
ML 347 × GM 8413	-2.19	-3.96	-12.88	-16.93	52.27	51.65	
ML 347 × KKM 3	0.46	-0.50	4.53	-6.55	5.42	2.22	
ML 347 × NP 36	0.81	0.10	0.55	-4.51	10.75	-1.45	
ML 347 × VGG 4	-2.84	-4.08	-9.76	-10.00	8.78	-0.28	
LM 567 × GM 8413	-3.20	-5.90	38.87	26.10	-11.94	-15.75	
LM 567 × KKM 3	2.40	2.34	-1.36	-21.91	34.48	32.10	
LM 567 × NP 36	2.72	2.40	1.79	-7.21	13.56	-3.11	
LM 567 × VGG 4	1.69	-0.61	13.34	-1.59	15.73	1.56	
SML $134 \times GM 8413$	-2.47	-4.01	-4.30	-17.27	26.84	13.36	
SML $134 \times KKM 3$	-1.06	-2.24	-31.47	-32.13	38.83	21.18	
SML 134 × NP 36	12.12 **	11.06 **	20.47	3.77	-42.38	-42.90	
SML $134 \times VGG 4$	-0.55	-1.58	-19.80	-27.51	55.44 *	51.75 *	
BL $862 \times GM8413$	-7.31 *	-7.35 *	2.13	-7.73	6.74	5.75	
BL $862 \times KKM 3$	-0.23	-2.91	-24.98	-29.27	27.76	23.25	
BL 862 × NP 36	3.31	0.78	0.13	-9.88	-14.15	-23.25	
BL $862 \times VGG 4$	-3.52	-4.01	-13.32	-17.91	18.56	41.55	
Hg 19/A × GM 8413	-4.31	-5.14	15.20	11.33	4.07	2.39	
Hg $19/A \times KKM 3$	0.87	-2.72	61.71 **	34.76	-17.58	-21.03	
Hg 19/A × NP 36	1.81	-1.56	35.49	31.48	19.71	7.69	
$Hg 19/A \times VGG 4$	-1.83	-3.21	-3.08	-4.71	-21.52	-27.24	
S.Em. ±	0.61	0.70	30.73	35.48	8.79	10.15	

^{*}Significant at P= 0.05 level **Significant at P= 0.01 level

Table.2 Overall heterotic status of hybrids in mungbean

Testers	GM 8413	KKM 3	NP 36	VGG 4	Hybrids with OHHS
Lines	(H)	(H)	(H)	(L)	J
LM 567 (L)	(240.5) H	(275) H	(259.5) H	(225.5) H	4
SML 134 (L)	(183) L	(240.5) H	(242) H	(142.5) L	2
ML 347 (L)	(149) L	(194) L	(221) H	(182.5) L	1
RMG 62 (H)	(190) L	(202) L	(229) H	(157) L	1
BL 866 (L)	(207) L	(113.5) L	(249) H	(283.5) H	2
Hg 19/A (L)	(260.5) H	(264) H	(303.5) H	(232.5) H	4
BL 862 (H)	(193.5) L	(239.5) H	(206) L	(241.5) H	2
Hybrids with OHHS	2	4	6	4	16

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(H): Over all high general combiner; (L):

Over all low general combiner

H: Over all high specific combiner; L:

Over all low specific combiner OHHS: Overall high heterotic status

Table.3 Distribution of heterotic crosses in relation to overall gca and sca status of parents and hybrids in mungbean

Parental	No. of crosses	No. of crosses with	No. of crosses	Conditional probability	Conditional probability of	
	under	high overall sca	with high	of given cross belonging	given cross belonging to high	
gca	category	status	heterotic status	to high sca status	heterotic status	
НхН	6	5	2	0.31	0.13	
HxL	2	1	1	0.06	0.06	
LxH	15	8	10	0.50	0.63	
LxL	5	2	3	0.13	0.19	

Only one cross Hg $19/A \times KKM \ 3 \ (61.71 \ \%)$ revealed positive significant heterosis over mid parent for SLA (cm²/g). Whereas, none of the crosses revealed significant heterosis over better parent.

Three crosses showed significant positive heterosis over mid parent for SCMR. viz., SML 134 \times VGG 4 (55.54 %) followed by BL 866 \times KKM 3 (53.67 %) and LM 567 \times GM 8413(8.05 %) BL 866 \times GM 8413(53.63 %). Whereas, only one cross SML 134 \times VGG 4 (51.75 %) revealed positive significant heterosis over better parent

Overall heterotic status of hybrids

In addition to determining the overall general combining ability status of parents and overall specific combining ability status of crosses, it is also important to determine the overall heterotic status of the hybrids across the characters.

Hence, the same method which was used for determining overall sca status of hybrids was also utilized to compute overall heterotic status of each cross and the results of the same are tabulated in table 2. It is evident from the table that, 16 out of 28 hybrids manifested high (H) overall heterotic status while the remaining 12 crosses expressed low (L) overall heterotic status across the traits.

As in the case of gca and sca, the magnitude and direction of heterosis varied considerably among the characters and therefore, it was difficult to decide the superiority of a hybrid over better parent. Hence, to decide whether a hybrid could be considered overall heterotic or not, the method proposed by Arunachalam and Bandyopadhyay (1979), with slight modification as by Mohan Rao (2001) was used. The results obtained clearly indicated high (H) overall heterotic status of 16 hybrids, while the remaining 12 hybrids had low (L)

heterotic status. The hybrids, Hg $19/A \times NP$ 36, BL $866 \times VGG$ 4, LM $567 \times KKM$ 3 had highest total score over the final norm.

Inter-relationship of overall gca status, sca status and heterotic status

Based on the overall gca effects, the crosses were classified as HH (both the parents of the cross with high overall gca status), HL (female parent with high and the male parent with low overall gca status), LH (female parent with low and the male parent with high overall gca status) and LL (both the parents of a hybrid with low overall gca status). The relationship between overall gca and sca status (Table 3) clearly indicated that both the parents with low overall gca status (L × L) also produced hybrids with high (H) overall sca status and the hybrids with H × H gca combination of parents expressed high (H) as well as low (L) overall sca status. Contrary to this, the hybrids involving high $(H) \times low(L)$ or low (L) × high (H) gca status expressed high (H) overall sca status in higher frequency in most of the crosses indicating major role of non-additive gene action.

When both overall gca status and overall heterotic status was considered, it clearly indicated that the hybrids involving $H \times H$ parental combination produced high (H) overall heterotic status, while, $L \times L$ parental combination produced low (L) overall heterotic status in the hybrids.

This suggested major role of additive gene action or additive \times additive interaction in such hybrids. However, the hybrids with H \times L or L \times H type of parental combination always produced hybrids with high (H) overall heterotic status, which indicated clearly the importance of non-additive gene action in the expression of heterosis. So these types of hybrid combinations are suitable for heterosis breeding. Whereas, the other

combinations of $H \times H$ or $L \times L$ can be utilized to get useful transgressive segregants in the subsequent segregating generations.

In total, the overall heterotic status depends on the gca status of parents, the best combiners always produced best hybrids.

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